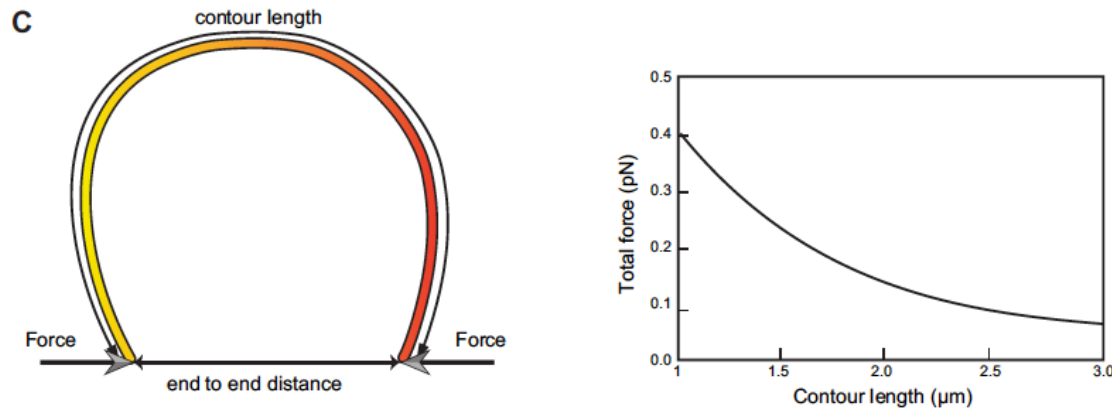
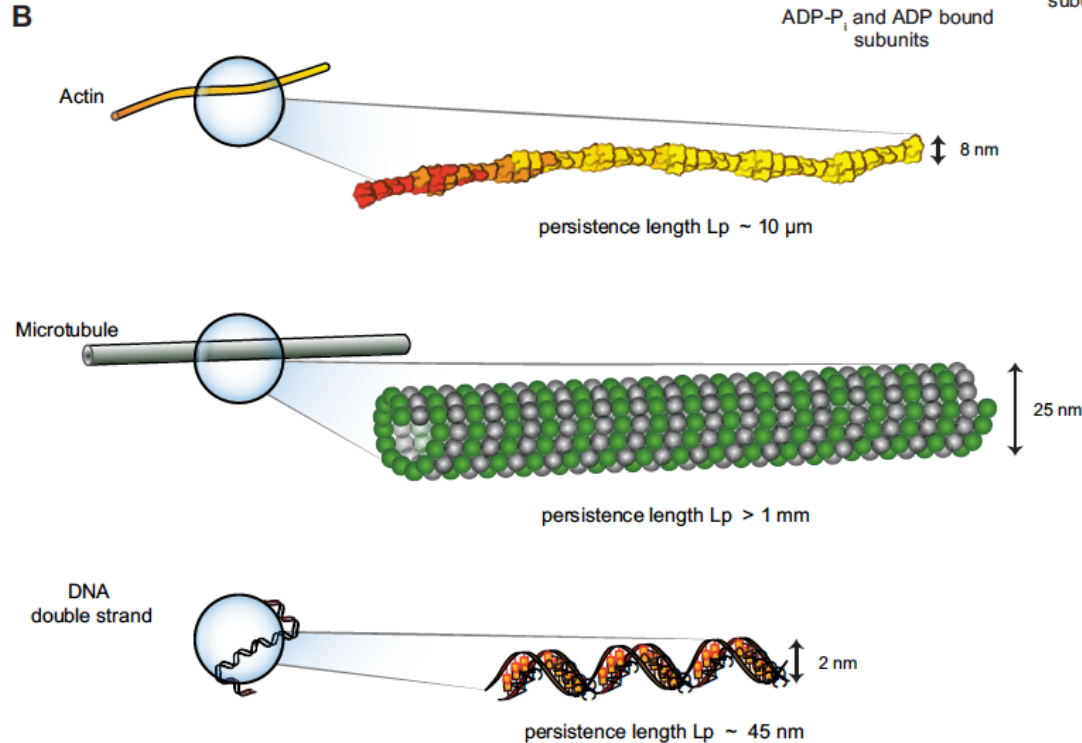
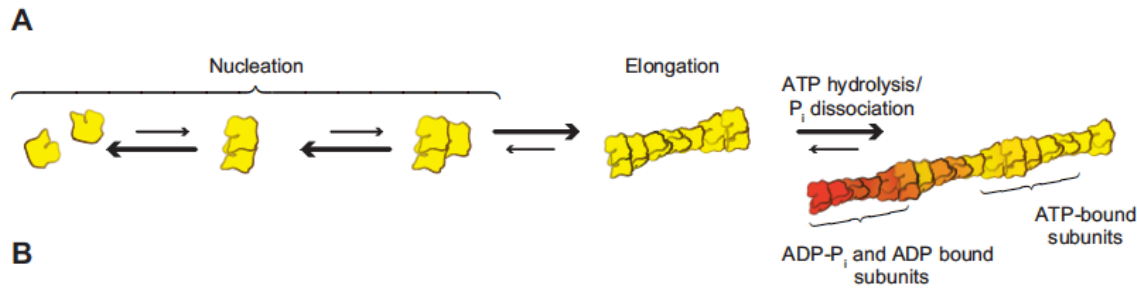


# *Single filament assembly and mechanics.*



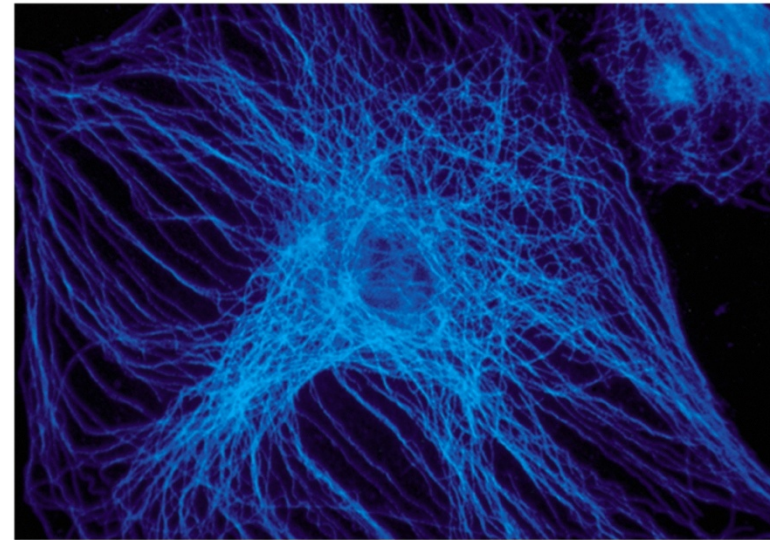
A: the kinetics of actin assembly.

B: persistence lengths of different cytoskeletal elements.

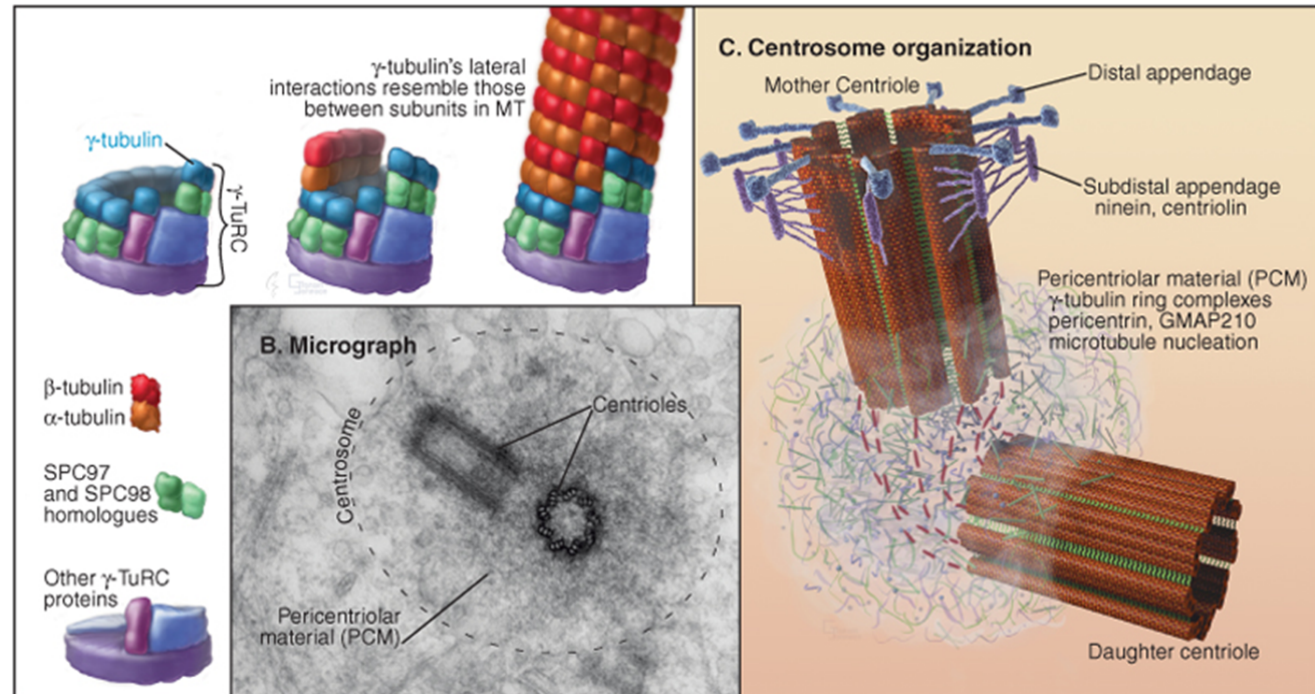
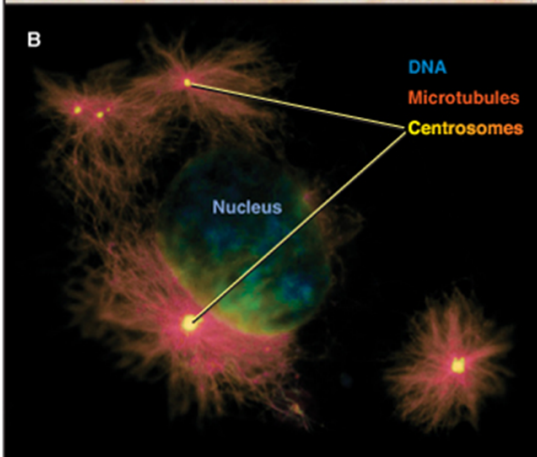
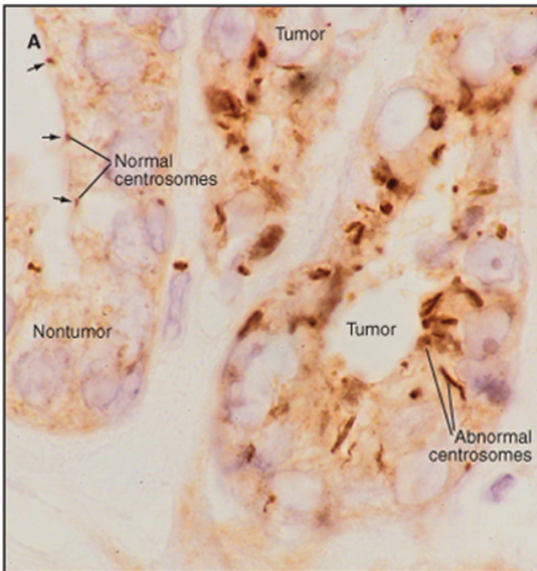
C: at the scale of the cell, actin filaments are almost straight structures, but they can nevertheless buckle under a load.

# Microtubules

Centrosomes are often abnormal in cancer cells.

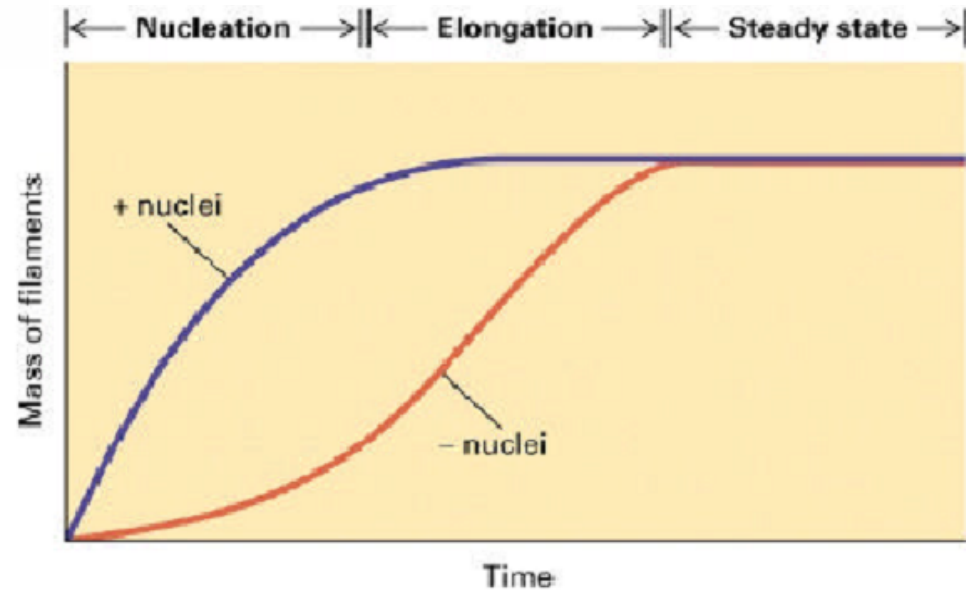
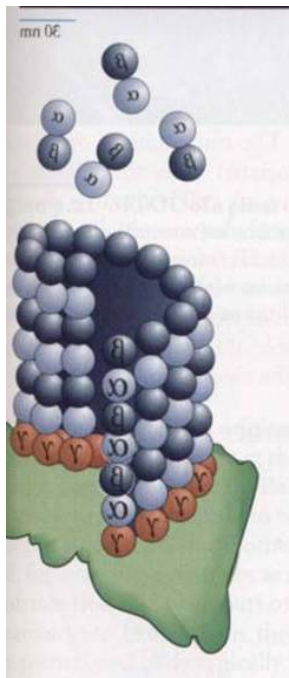
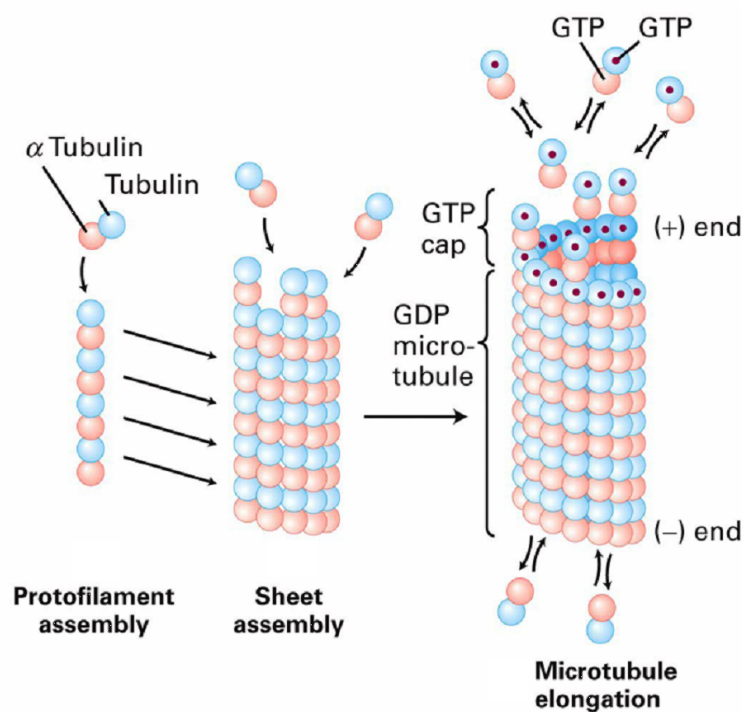


The centrosome is the primary microtubule nucleation site in most cells





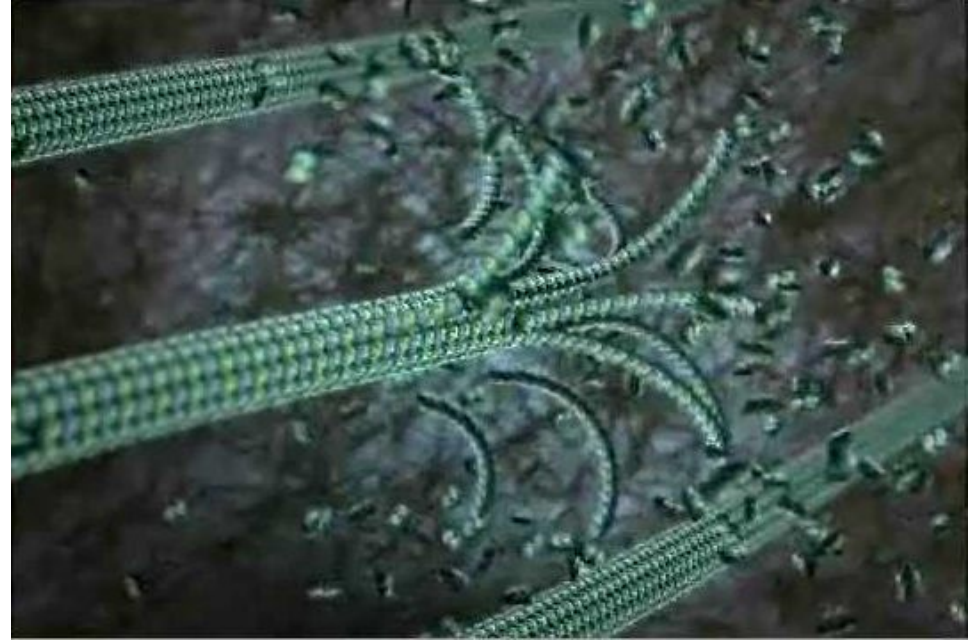
# *Tubulin assembly*



**$\gamma$ -tubulin at initiates synthesis at one end (-) (green).**

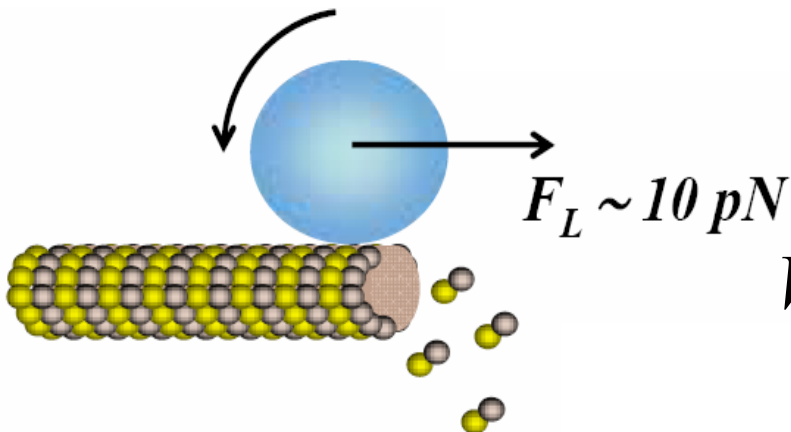
# Depolymerization

The elasticity of the protofilaments that curve outward at the disassembling plus end drives a sliding collar on the kinetochore toward the minus end (*a power stroke mechanism*).

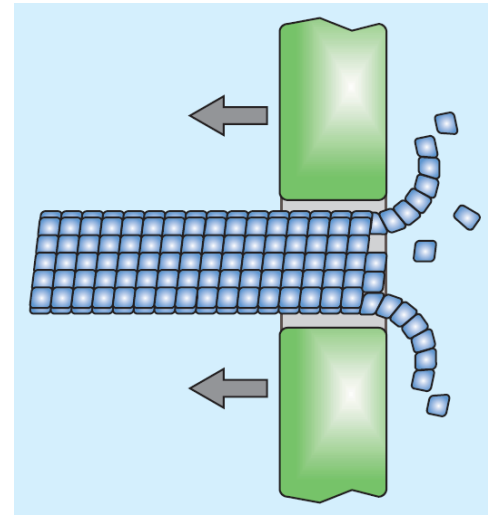


$$F_L \cdot \delta = k_B T$$

$$\delta = \frac{k_B T}{F_L}$$



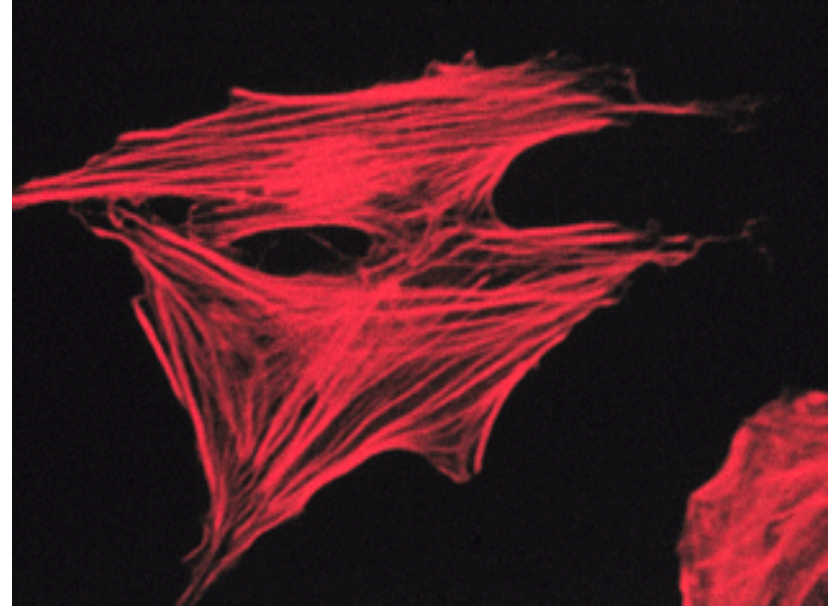
$$V_p = k_{off} \delta = \frac{k_{off} k_B T}{F_L}$$





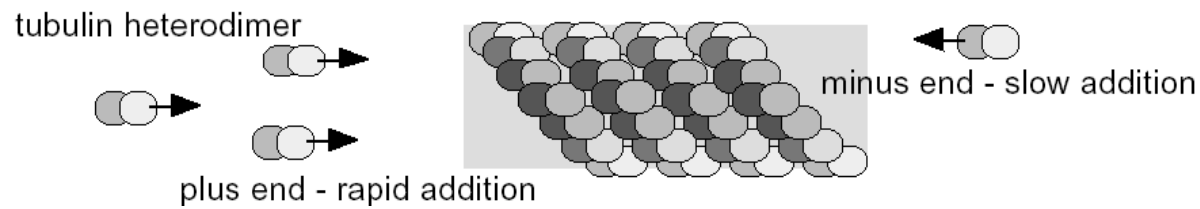
*Tubulin half life is  
nearly a full day*

*The half life of a given  
microtubule may be  
only 10 min.*

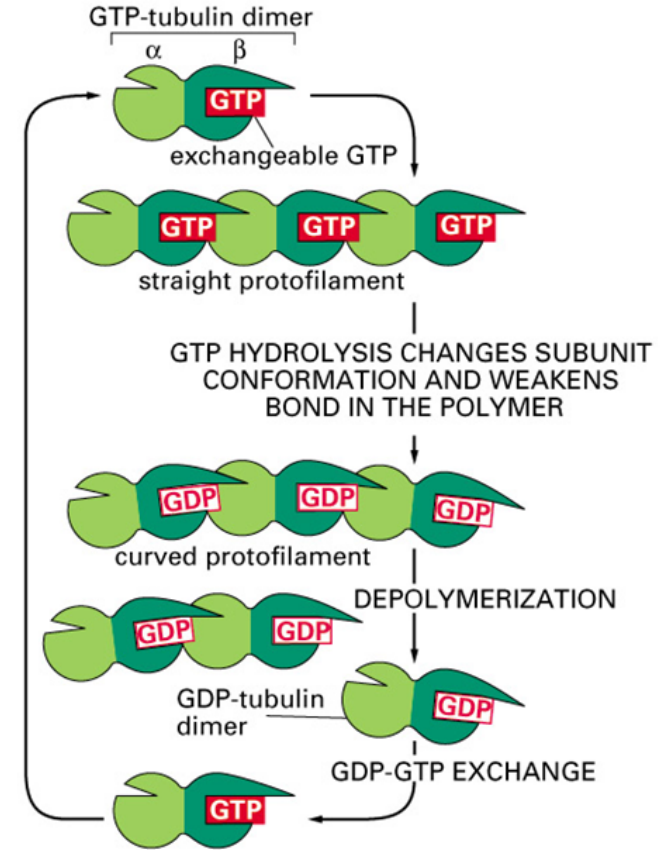
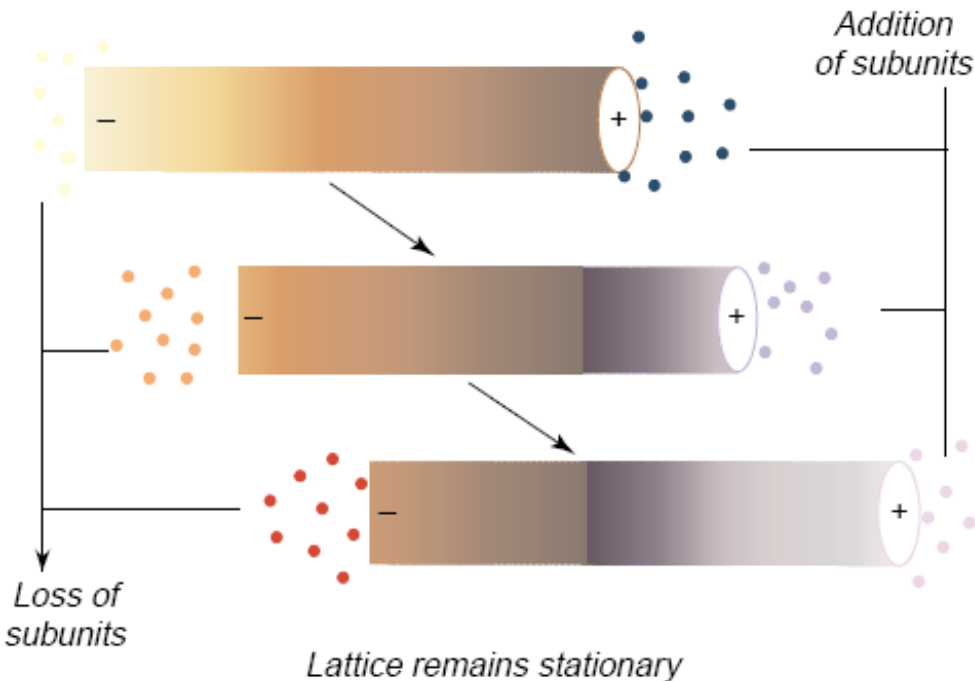


- ✚ A static microtubule grow in the presence of a non-hydrolysable GTP analogue - a tube has little tensions.*
- ✚ A dynamic microtubule grow in the presence of GTP is a tube ready to crack.*

The tubulin concentration in cells is on the order of 20  $\mu\text{M}$



- Subunits on the fast end contain GTP.
- The slow end has an GDP containing subunit and depolymerizes.



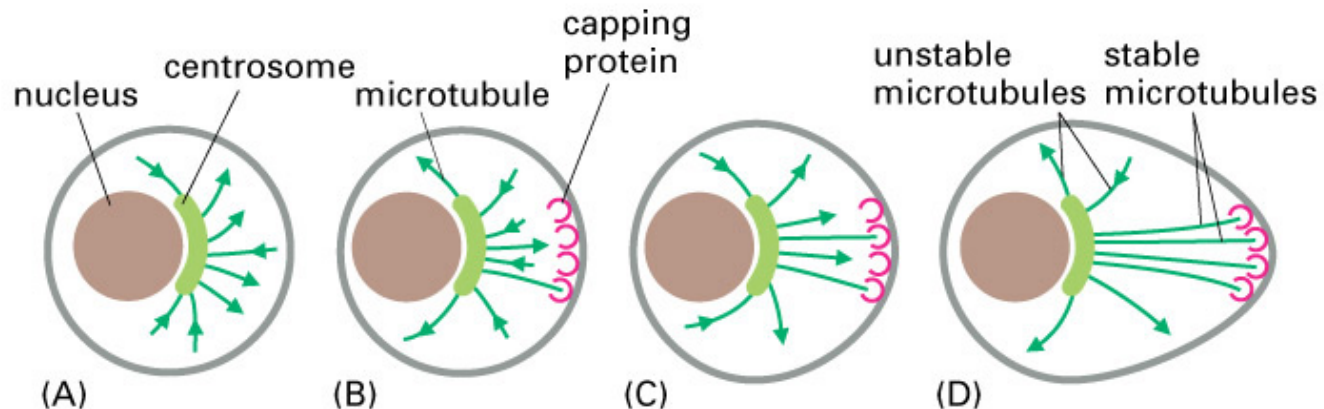
## *Treadmilling*

*Treadmilling does not involve actual movement of the microtubule lattice.*

# *Dynamic instability: significance*

- Allows 'random' searches
- Allows rapid re-arrangements of MTs
- Makes 'selective stabilization' (at tip) possible
- Can be locally regulated, to drive MTs into or out of an area
- Microtubule dynamics allow the cell to quickly reorganize the network when building a mitotic spindle
- Dynamics also allow microtubules to probe the cytoplasm for specific objects and sites on the plasma membrane - search and capture

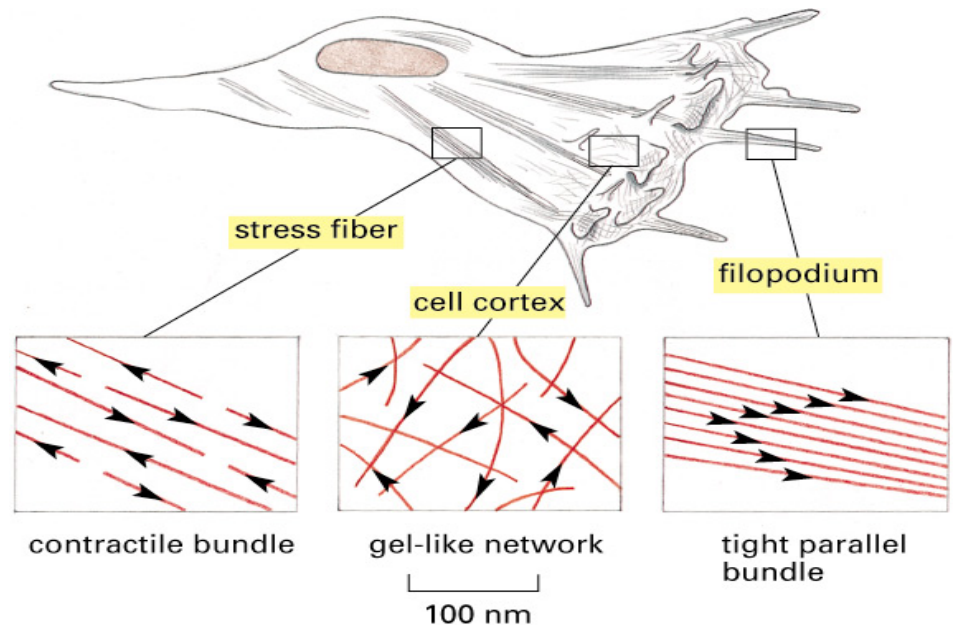
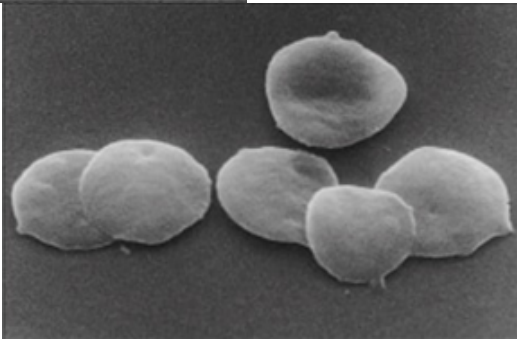
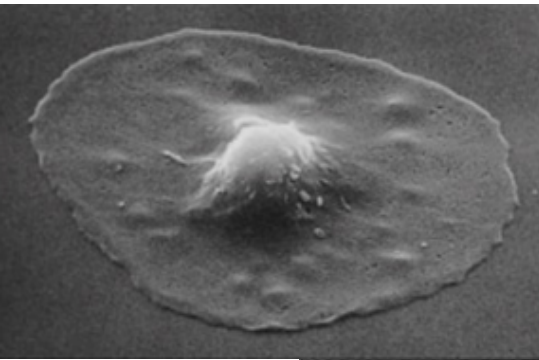
*Search & capture  
during cell  
polarization*

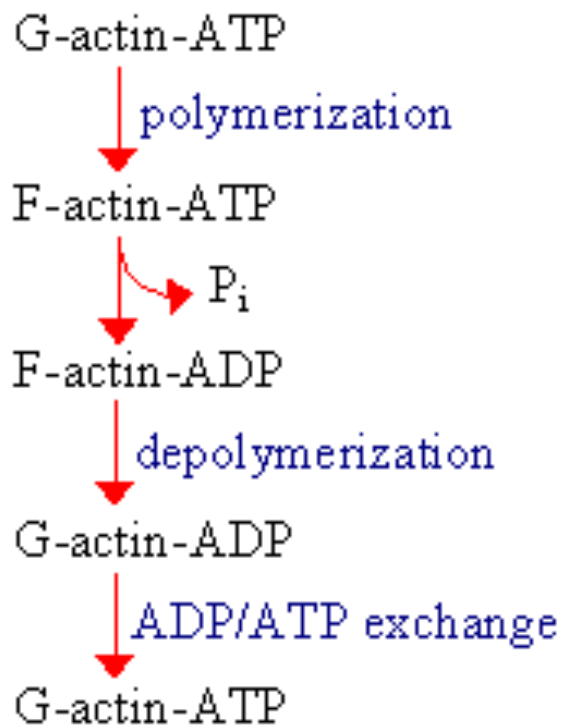




# *Actin based cell movement*

- Wound healing – epidermal cells.
- Immune response – leukocytes – migrate to sites of infection.
- Development – neural crest cells; neuronal process extension.
- Cancer cell metastasis – malignancy determinant.

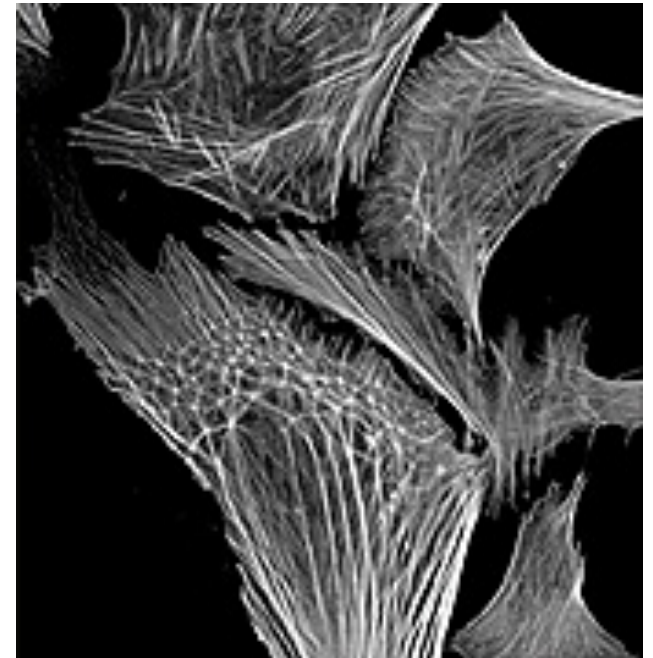




*During growth, the energy supplied by ATP/GTP hydrolysis is stored in the lattice as mechanical strain.*

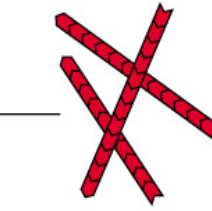
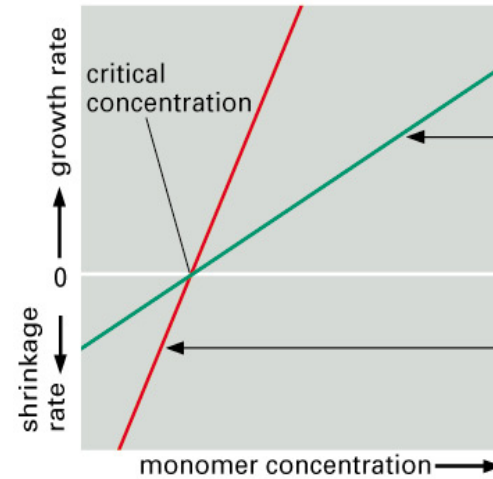
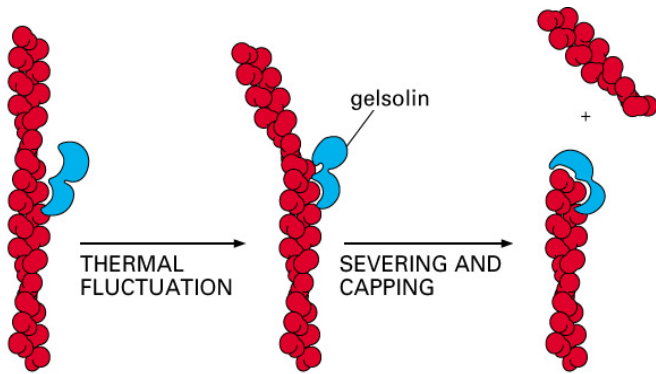
*This strain powers the fast shortening of disassembling microtubules.*

*Fibroblasts-actin filaments*



*Dynamic instability and treadmilling are phenomena that require energy dissipation, and which could not emerge from a pure self-assembly process.*

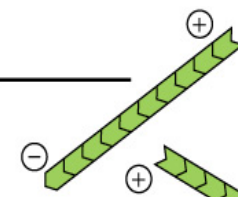
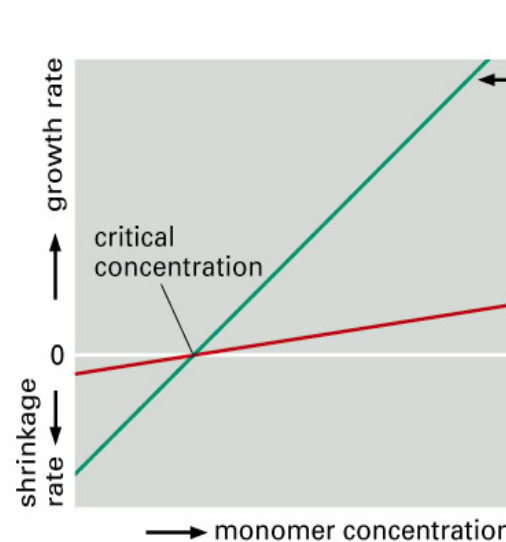
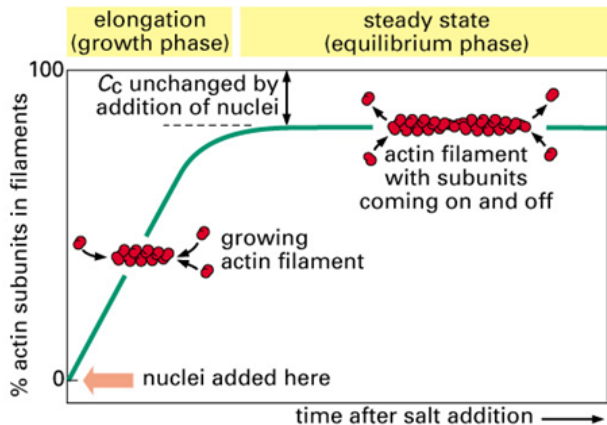
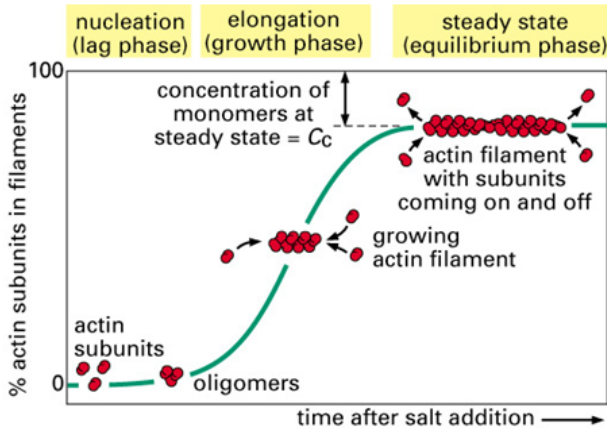
# Regulation of polymerization rate



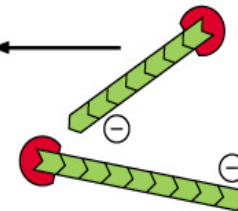
in uncapped population, actin filaments grow and shrink relatively slowly



in capped population, actin filaments grow and shrink more rapidly



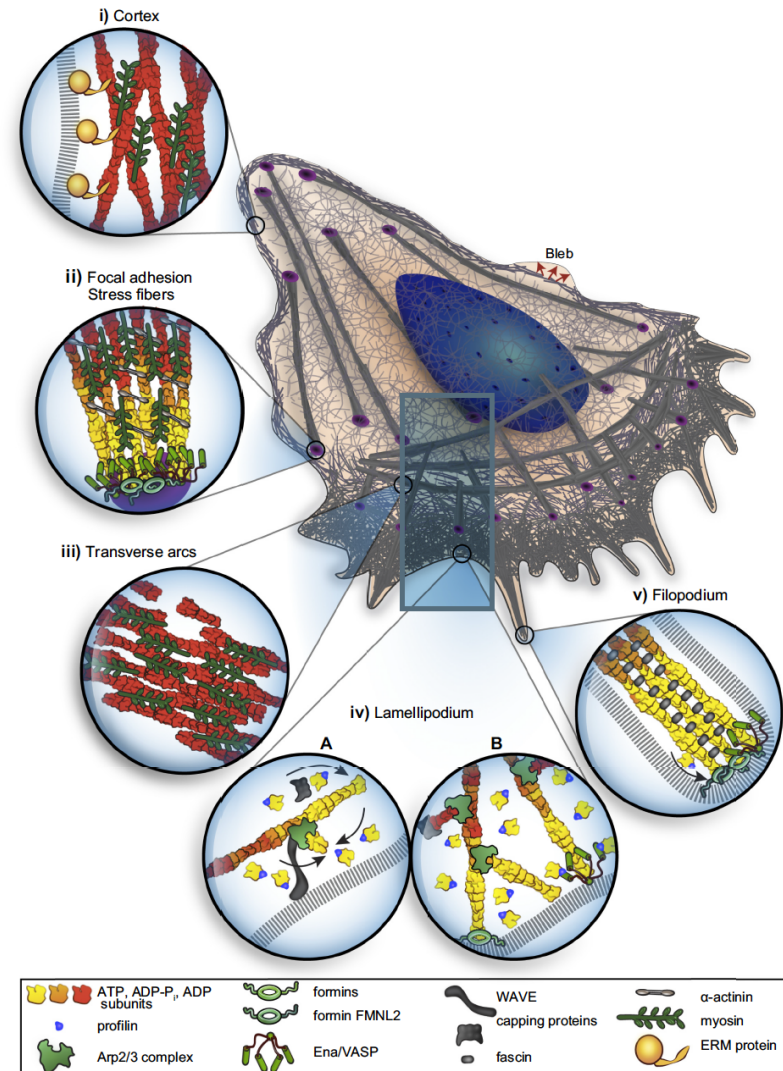
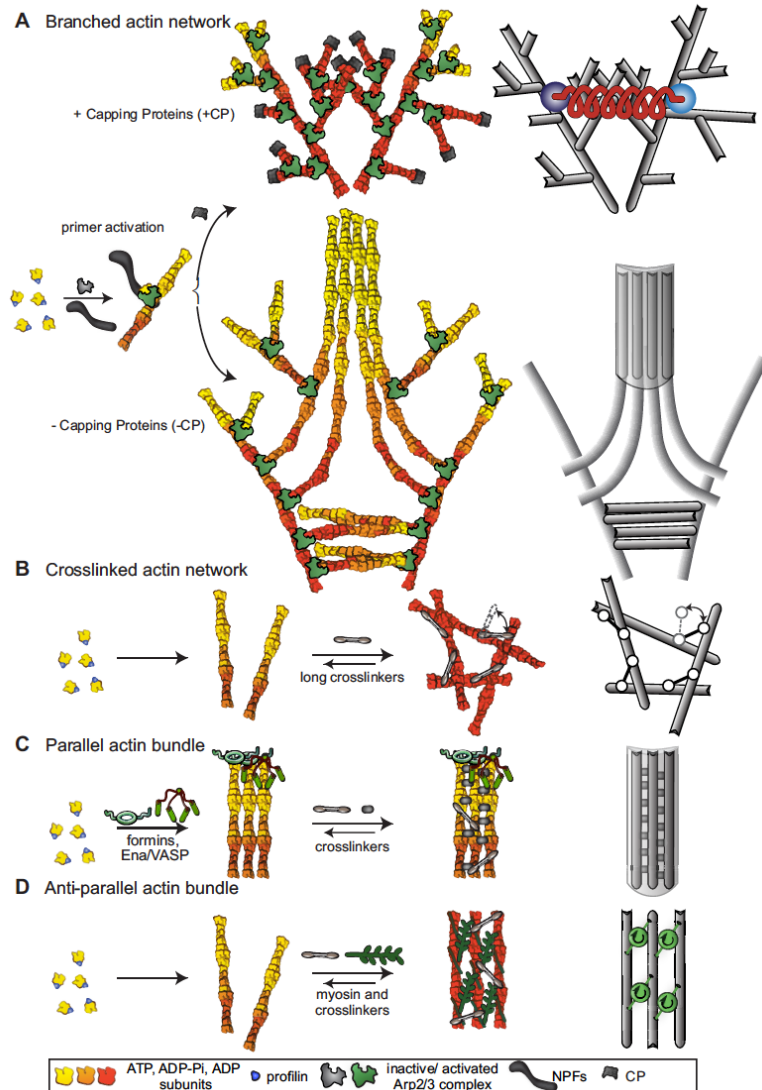
uncapped population of filaments: growth at plus and minus ends

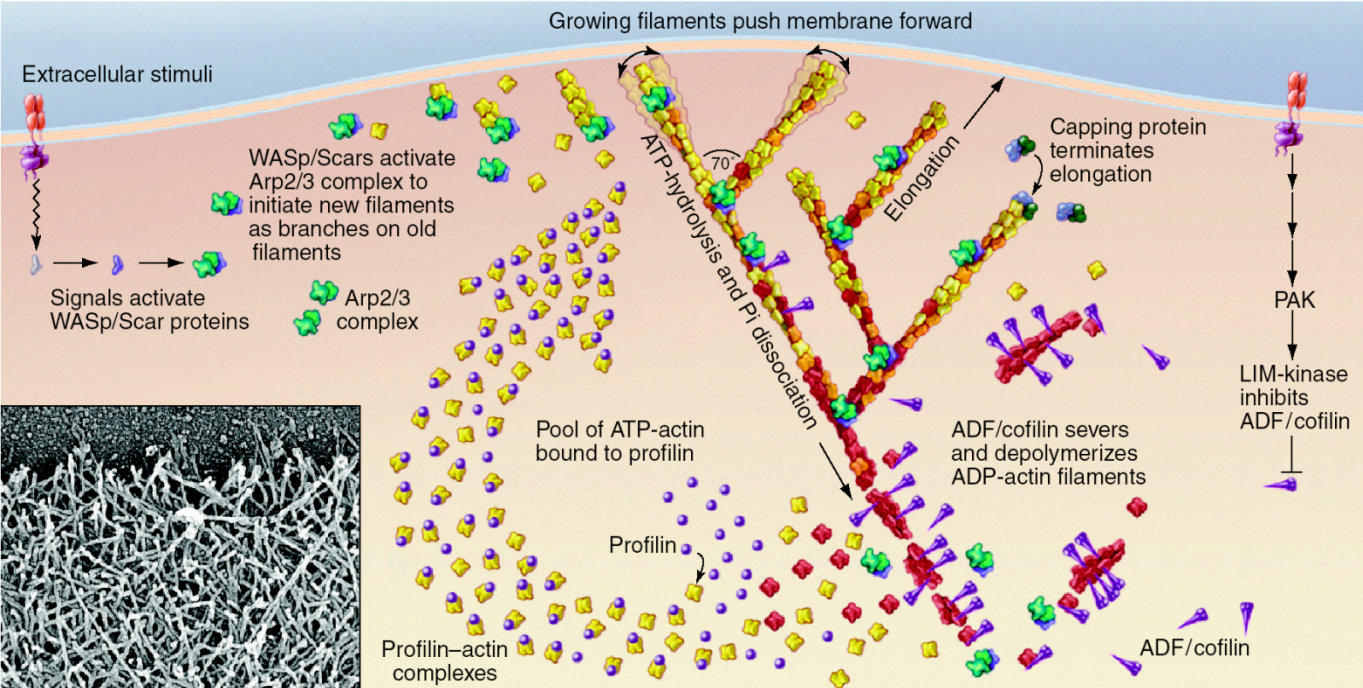


capped population of filaments: growth at minus end only

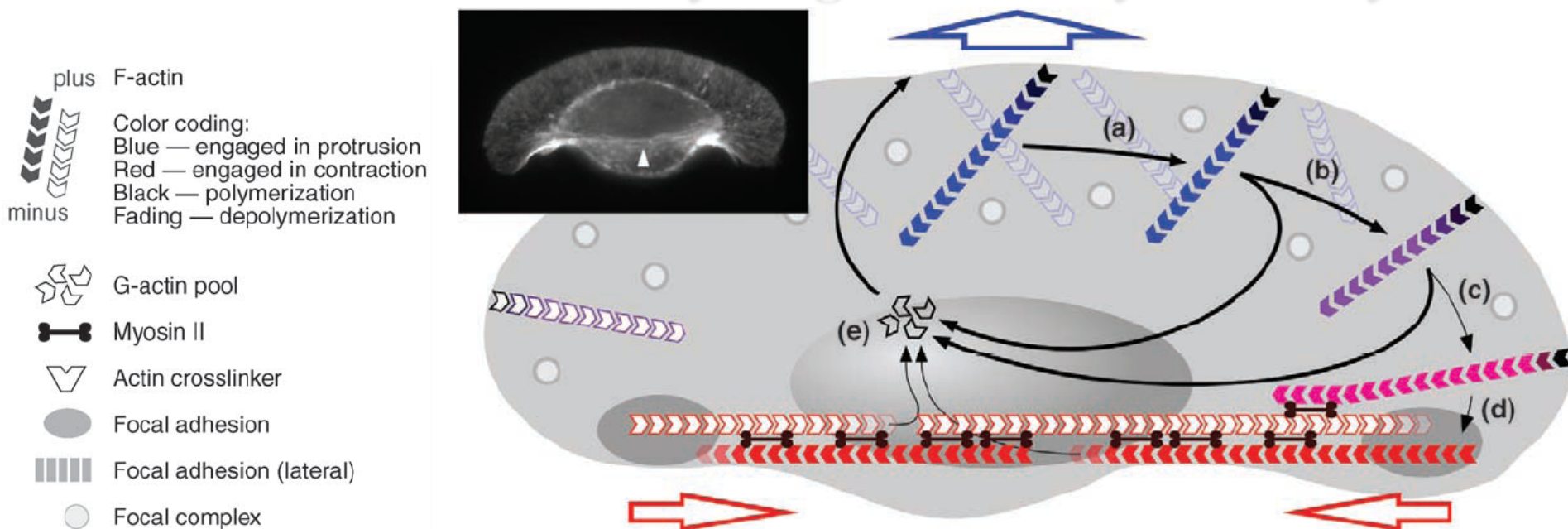


# *Distinct actin filament organizations and their mechanical description.*





## *Filament recycling in keratocyte motility.*

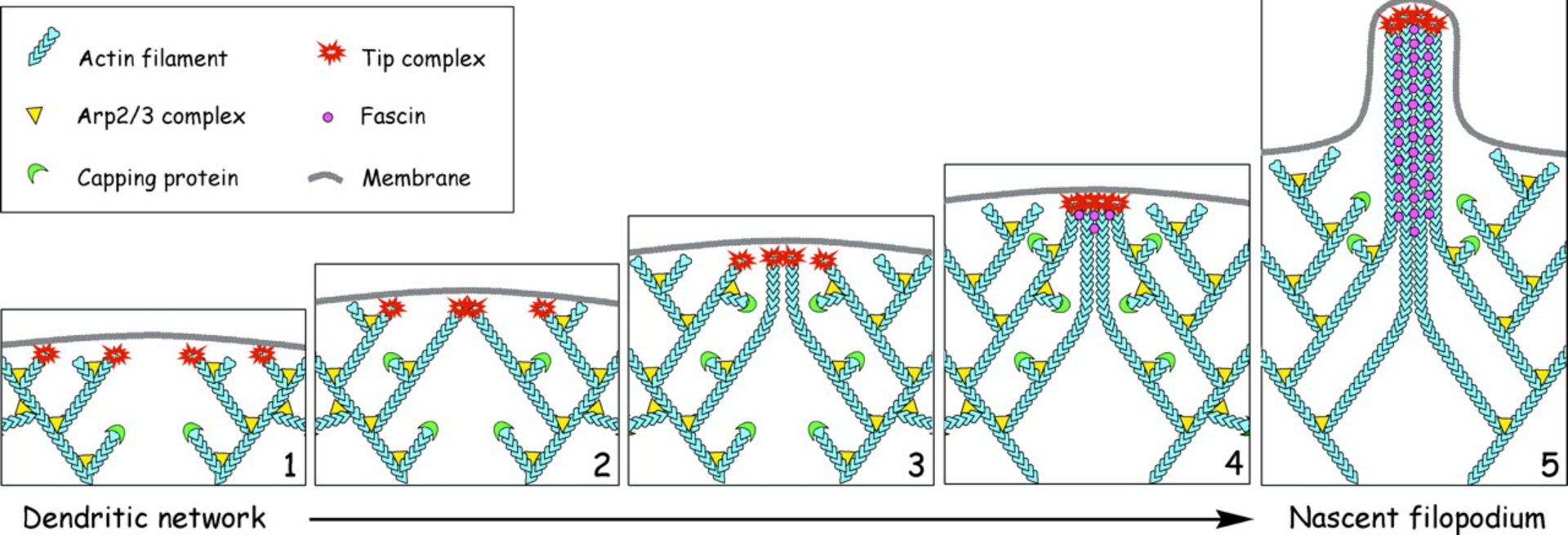
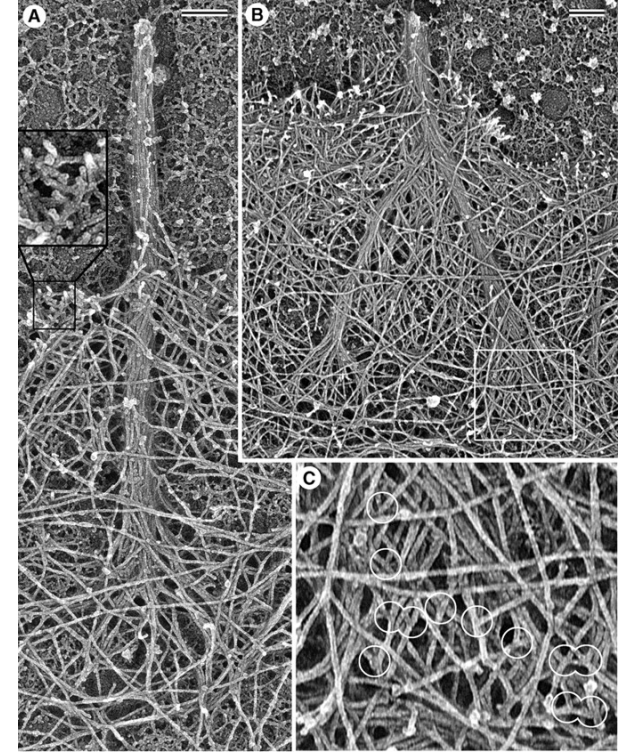




*Filopodia are composed of long, unbranched actin filaments*

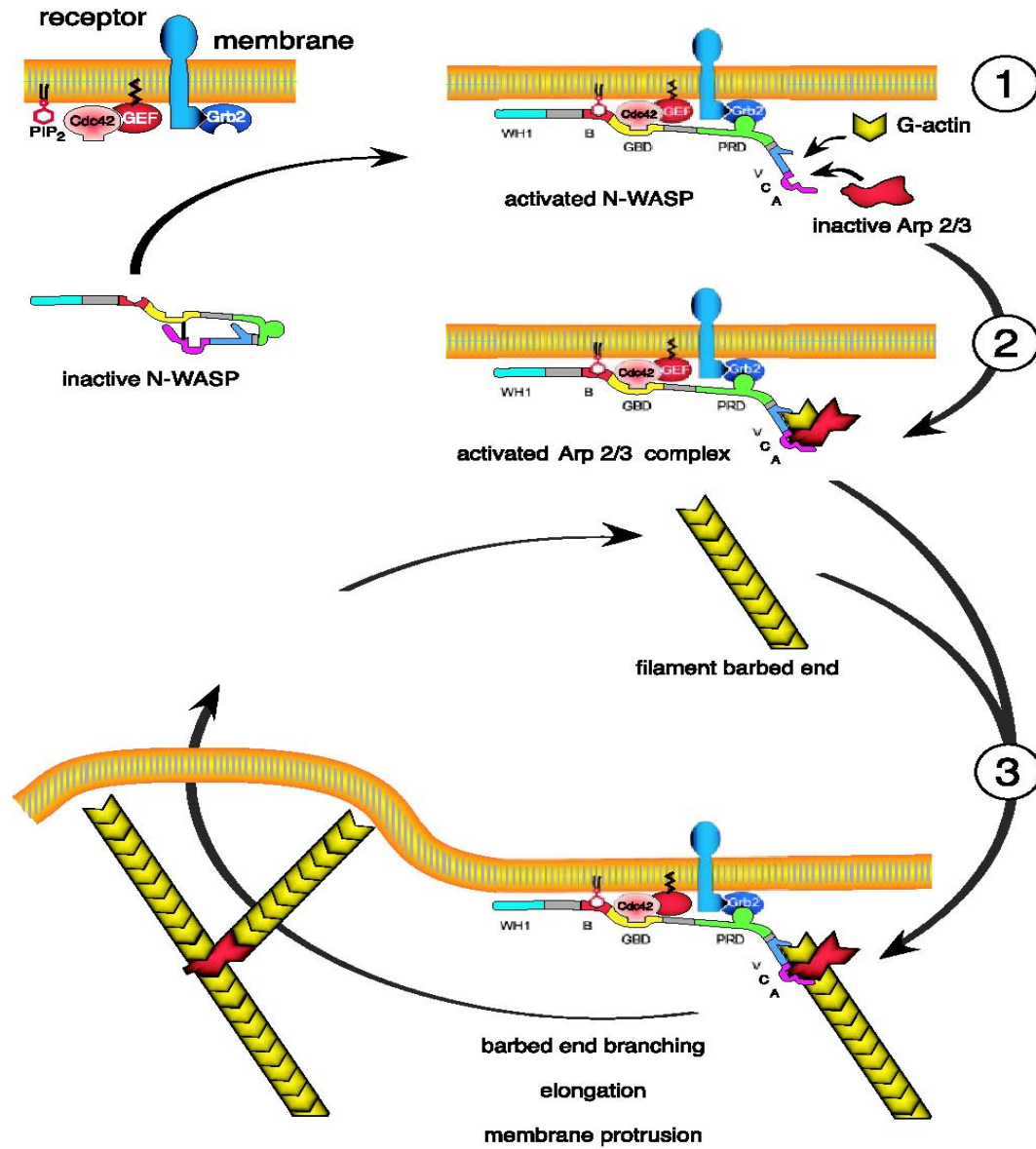
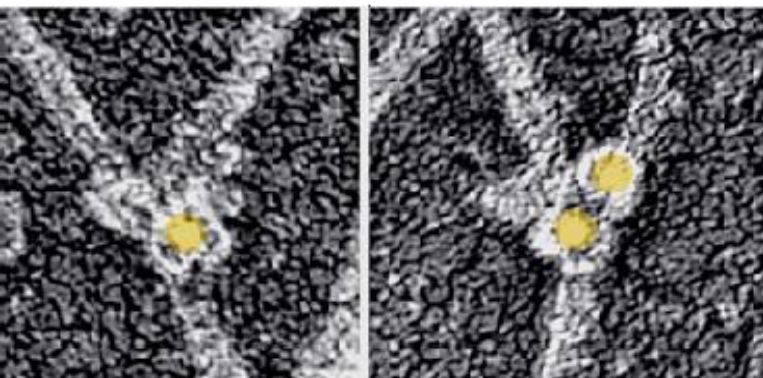
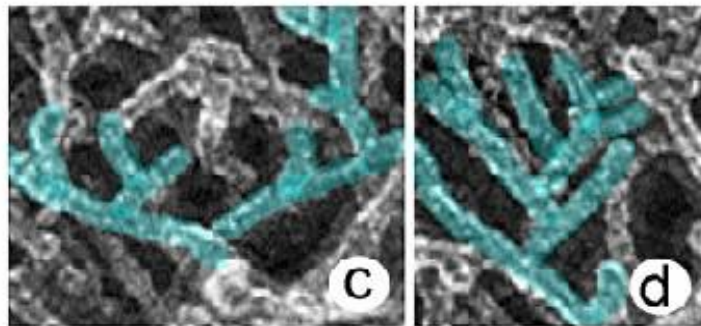
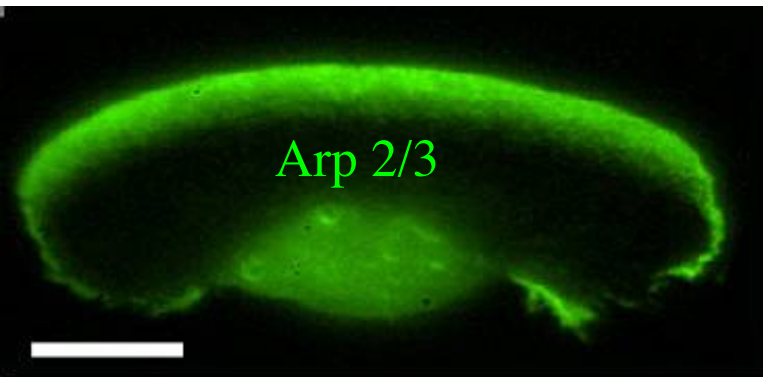
# *Generating movement*

## *Actin dynamics in filopodia*





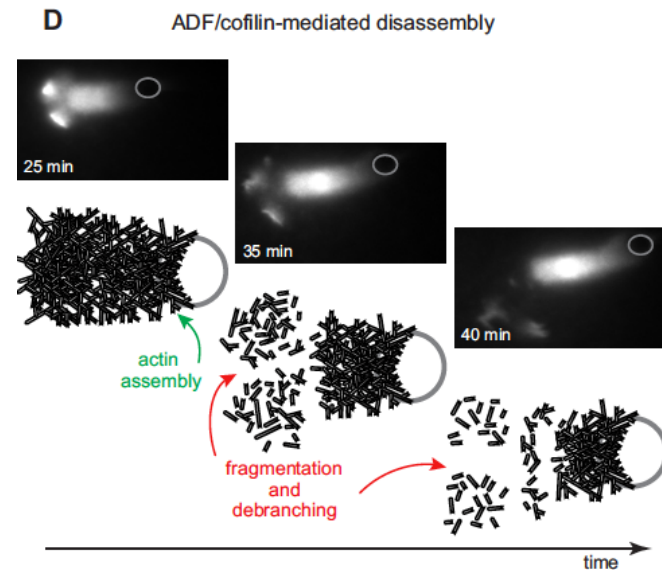
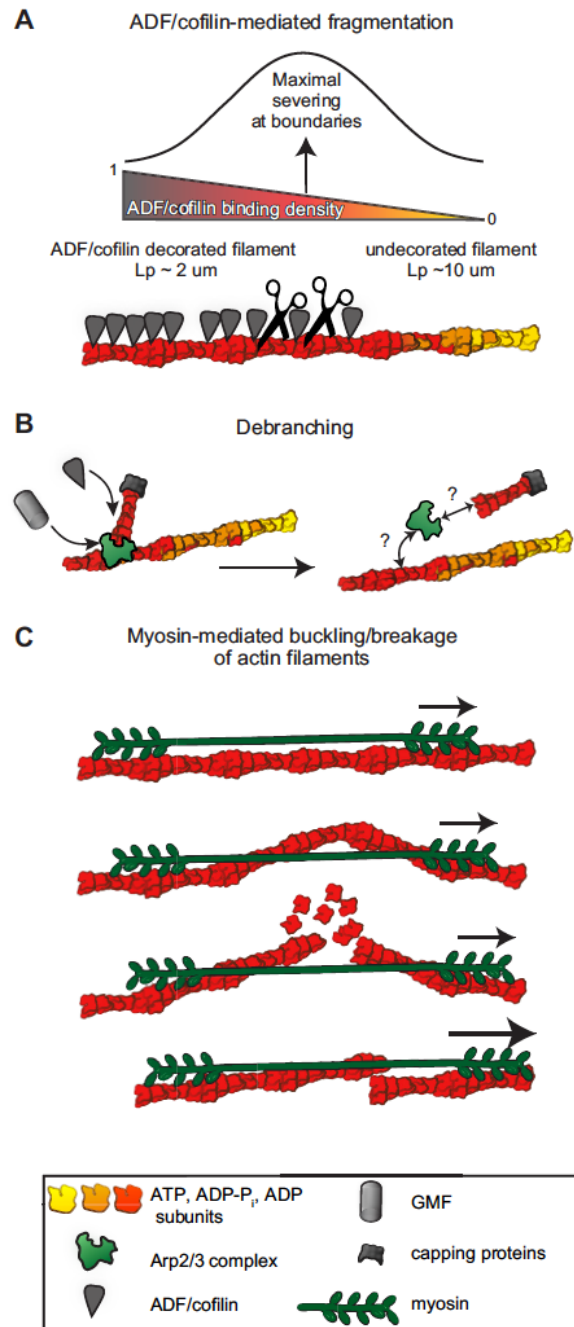
# *Dynamics of actin polymerization at the leading edge*



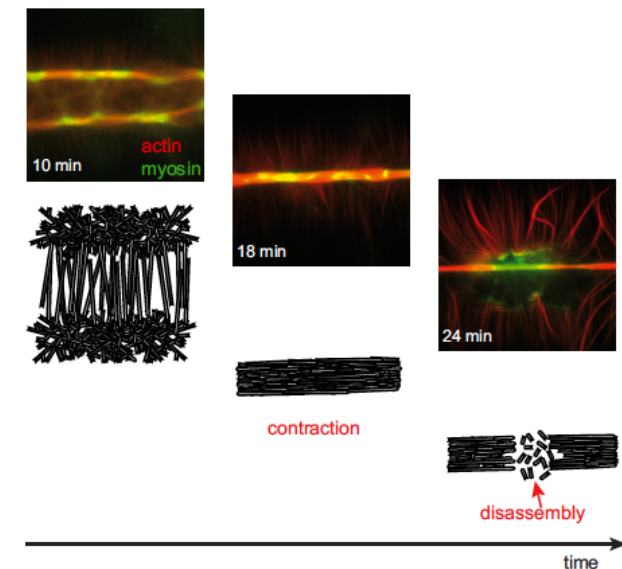
## Microscopic disassembly of actin filaments

# Remodeling and disassembly of dynamic actin structures.

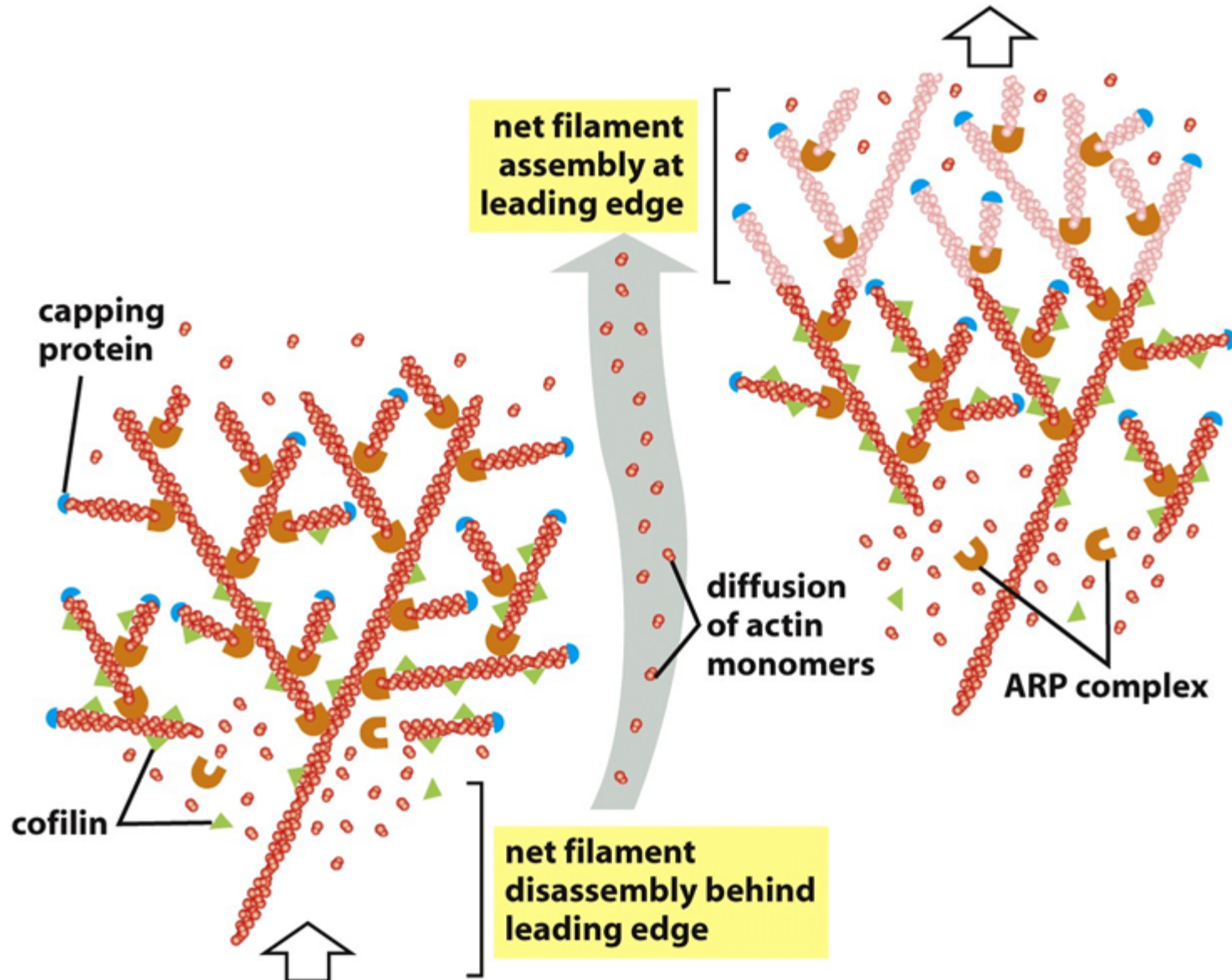
## Macroscopic disassembly of actin networks



**E** Myosin-mediated disassembly

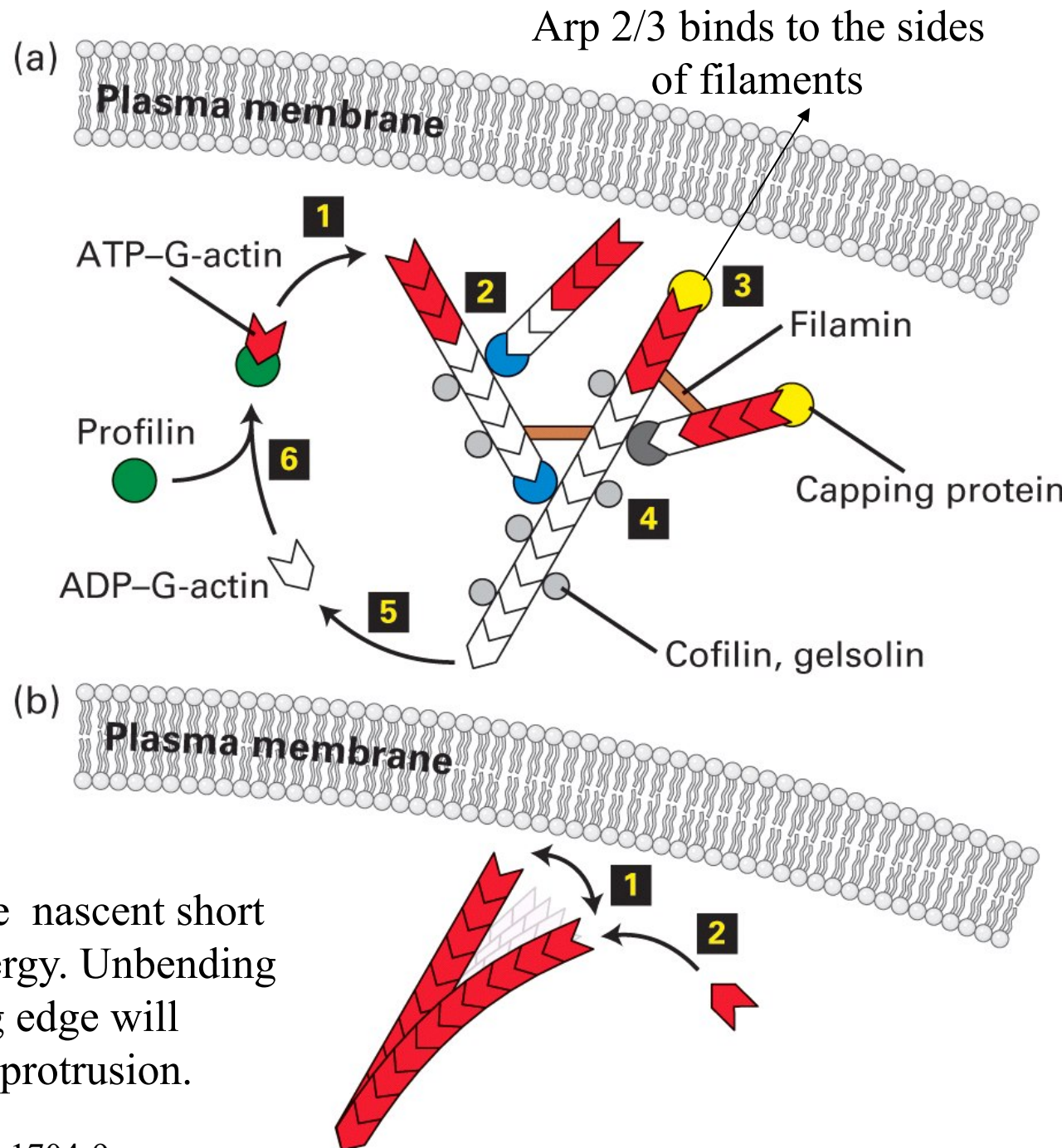


# *Polymerization based movement*



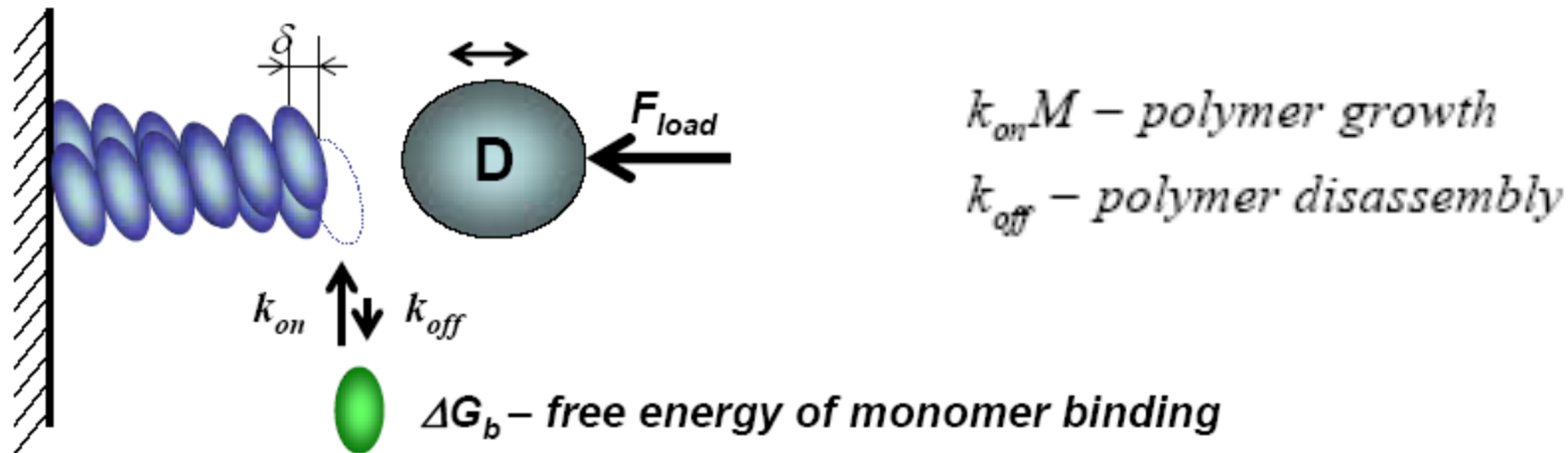


# ***“Elastic Brownian ratchet”***



The thermal energy bends the nascent short filaments, storing elastic energy. Unbending of the end against the leading edge will provide the driving force for protrusion.

# Polymerization motors – ratchet mechanism



$$\Delta G_{tubulin} = 5-10 k_B T$$

*The motor does not directly drive the load, but simply rectifies its Brownian diffusion.*

$$V_p = \delta \left( k_{on} M^* \exp\left(-\frac{F_L \delta}{k_B T}\right) - k_{off} \right)$$

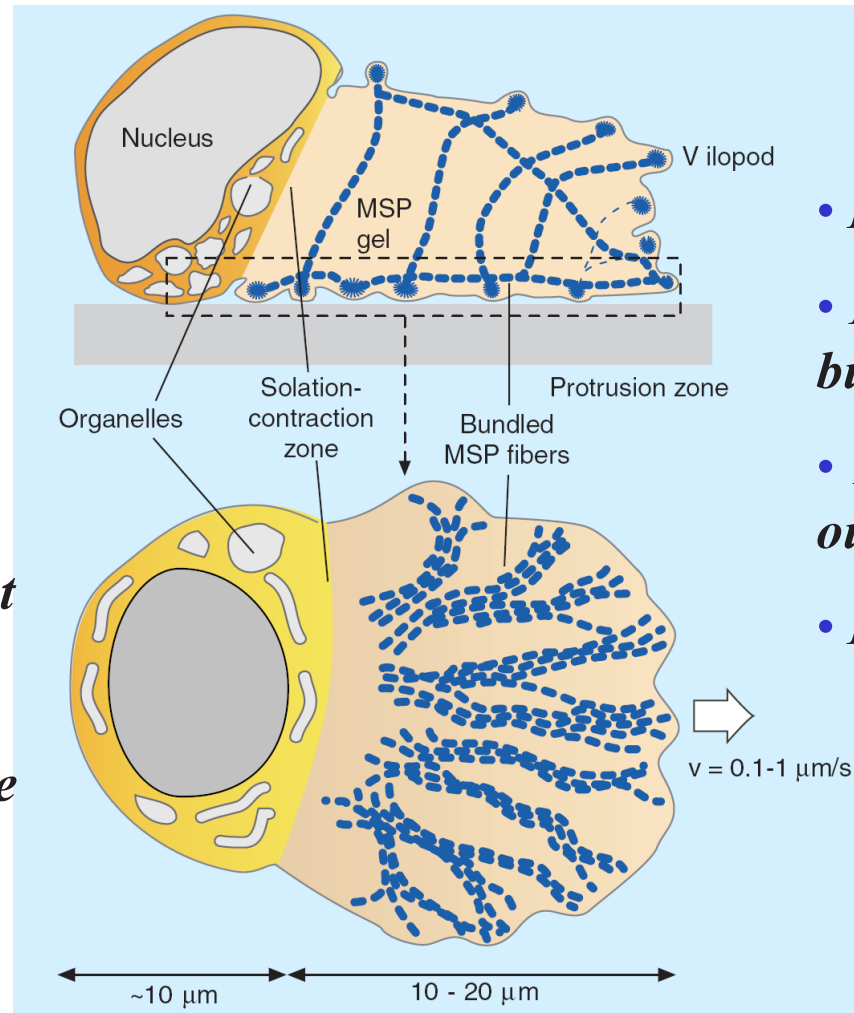
$$F_{stall} = \left( \frac{k_B T}{\delta} \right) * \ln \left( \frac{k_{on} M}{k_{off}} \right)$$

*Then, characteristic stall force ( $V=0$ ) is:*

# *The directional movement*

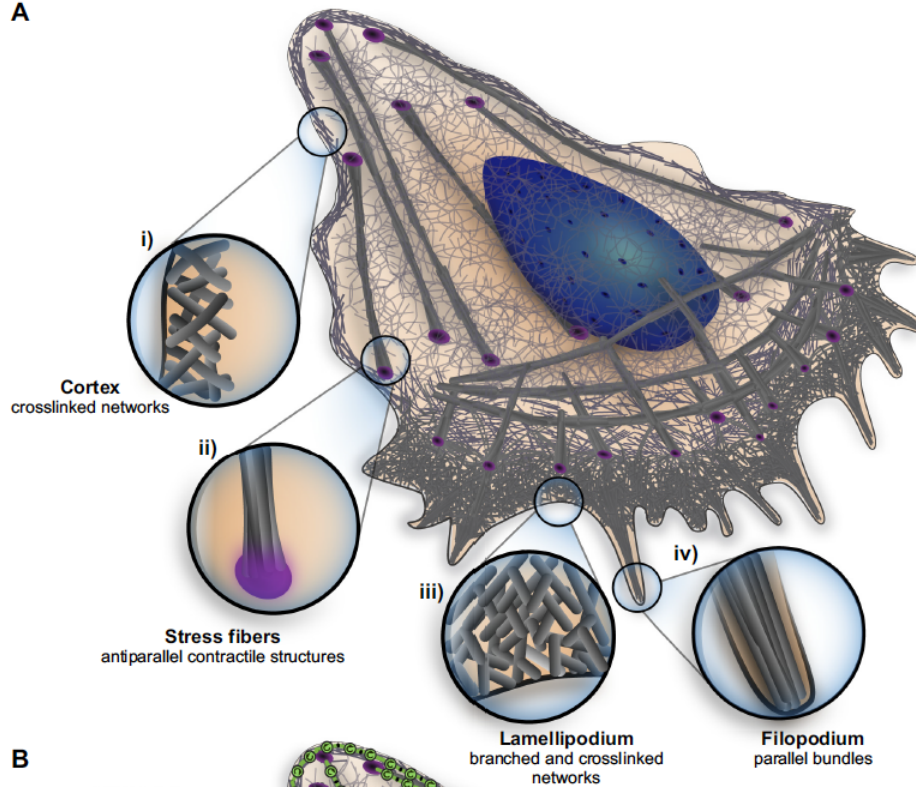
*The perinuclear mitochondria generate an anterior–posterior pH gradient which regulates gelation and solation of the gel in the lamellipod.*

- *Low pH.*
- *The interfilament interactions weaken.*
- *The filaments unbundle.*
- *Because the cell front adheres to the substratum, this provides the contractile force to pull the cell body forward.*



- *High pH.*
- *Filaments grow and bundle into thick fibers.*
- *The cell front is pushed out.*
- *Elastic energy is stored.*

A



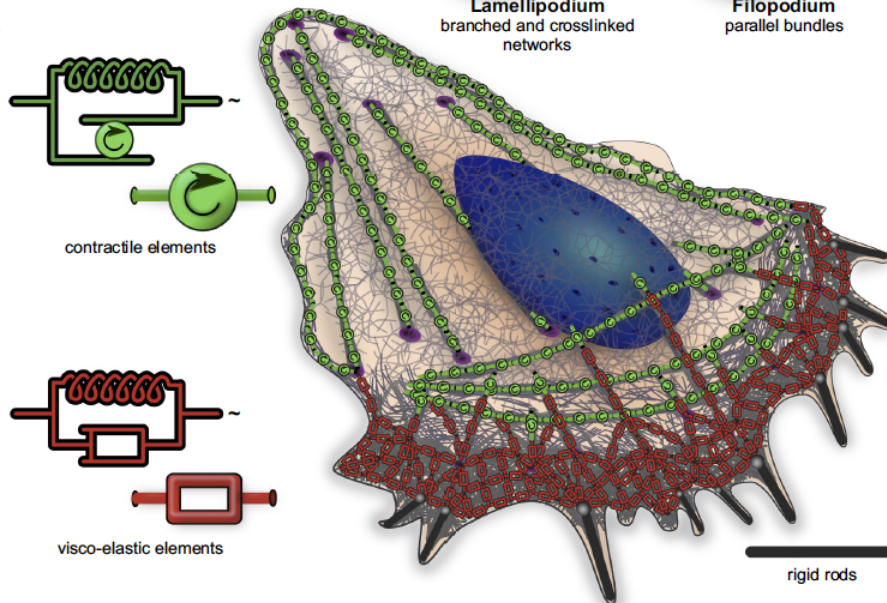
## *Overlay of actin architecture and mechanics in the moving cell.*

A: schematic representation of the cell with the different architectures.

B: overlay of the actin architecture and its mechanical profile.

*The red rectangles are the shock absorbers (dashpots) that represent the actin network, while the green circles are active springs due to myosin motor activity.*

B





# *Listeria utilizes the power of actin polymerization for intracellular movement*

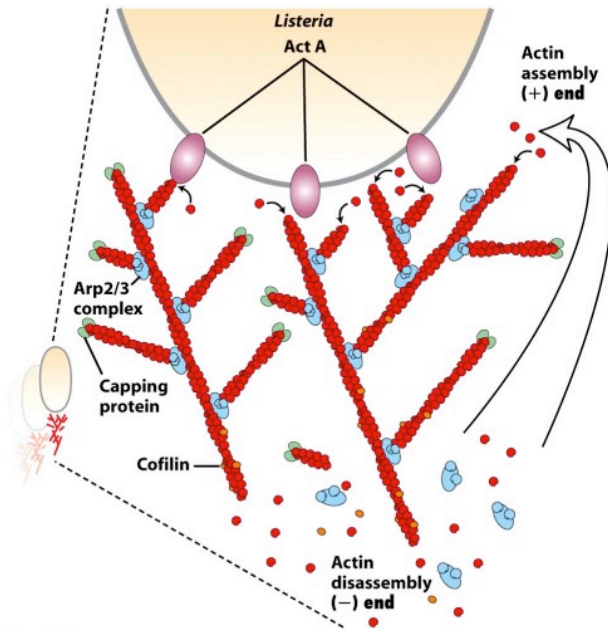


Figure 17-17c  
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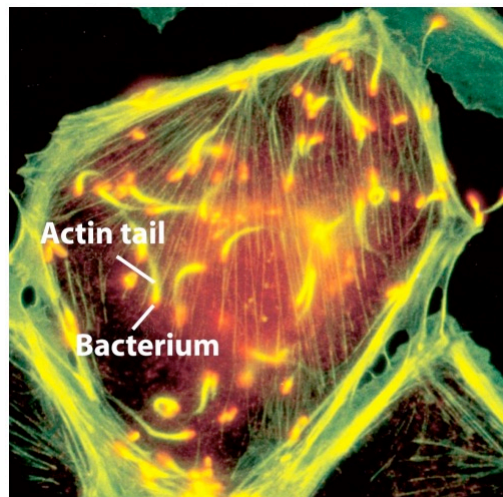
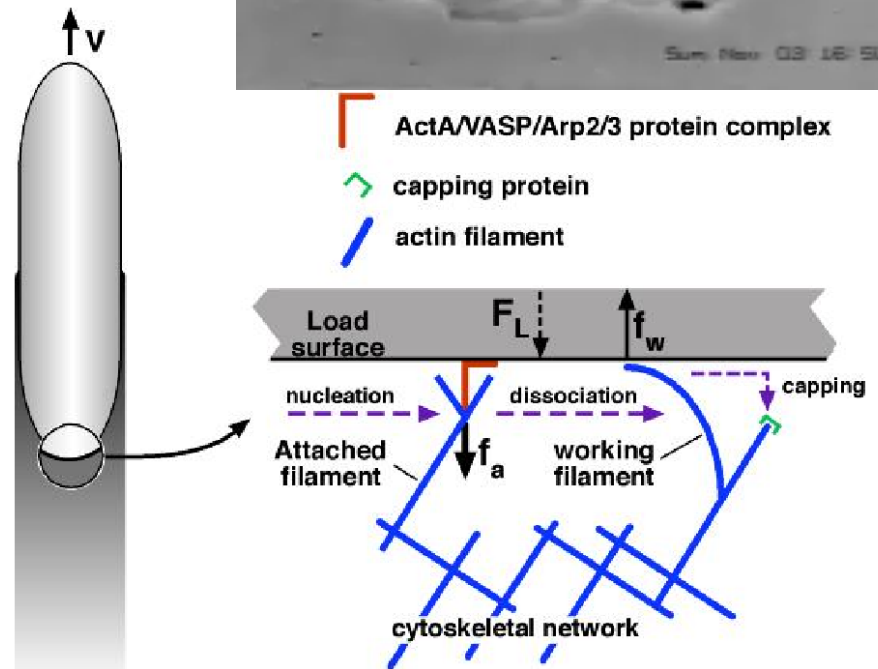
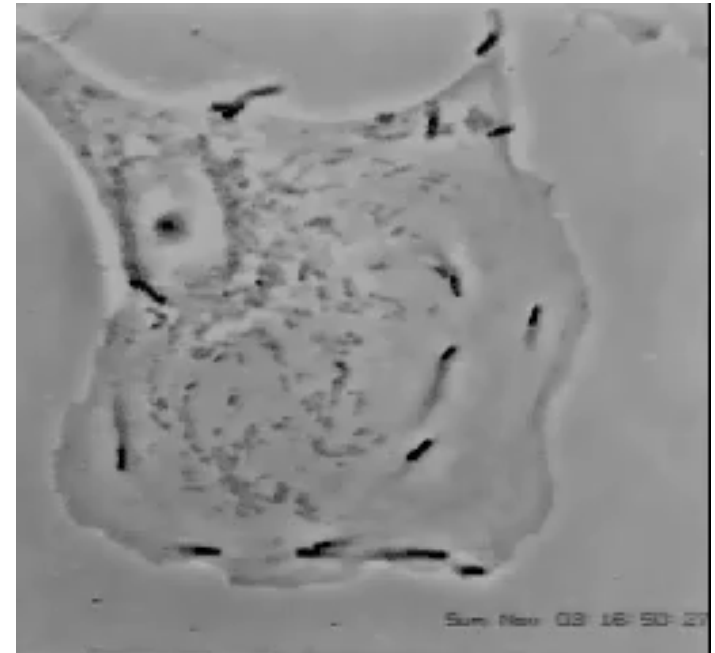
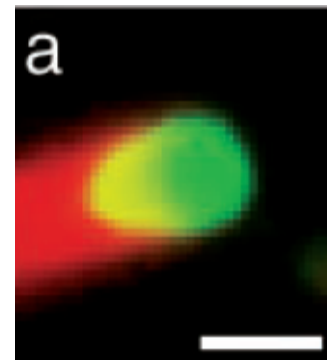
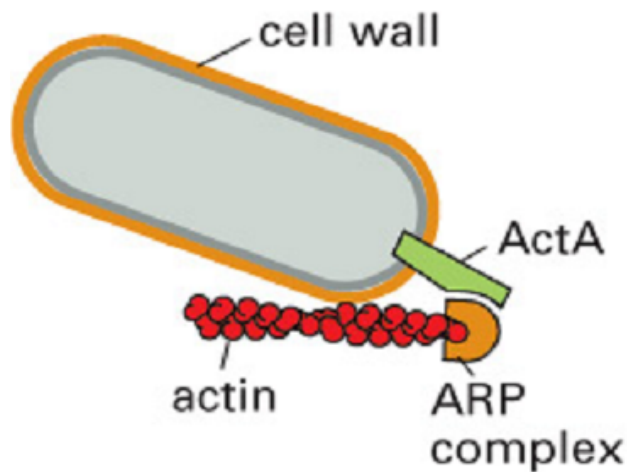
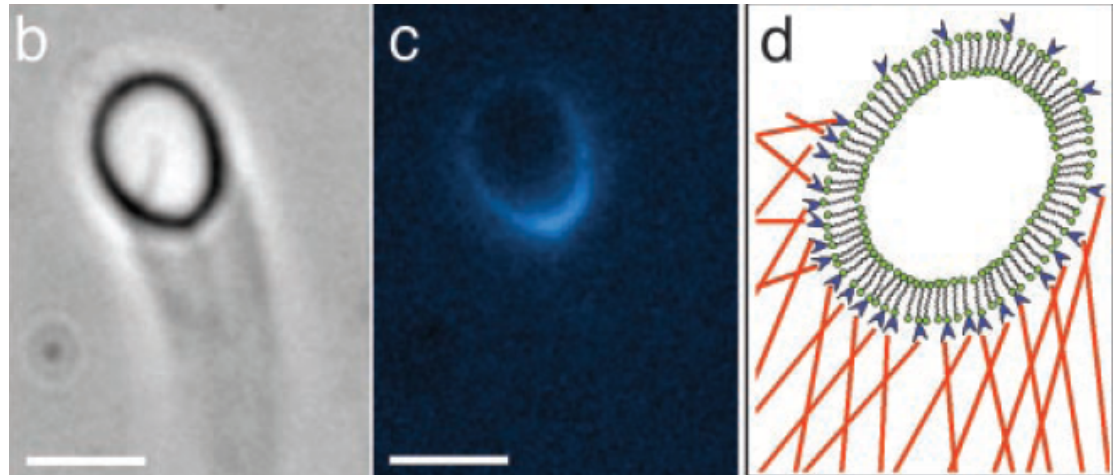
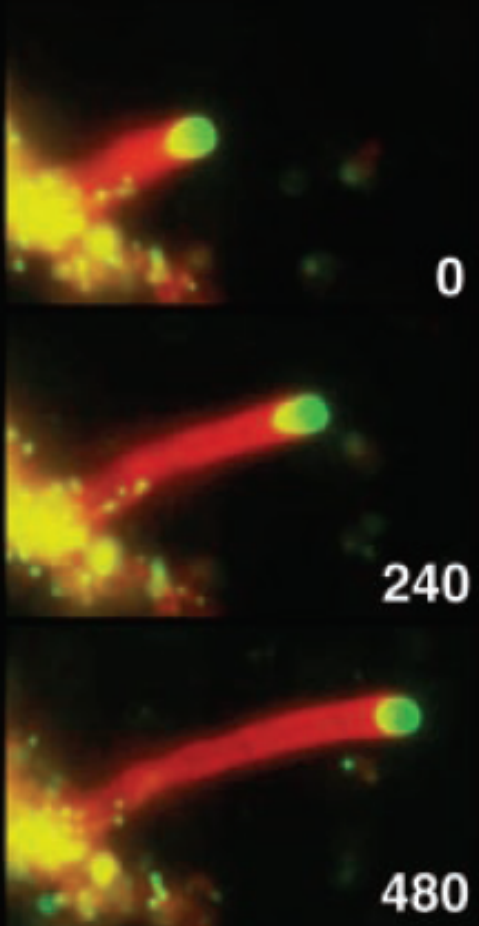


Figure 17-17a  
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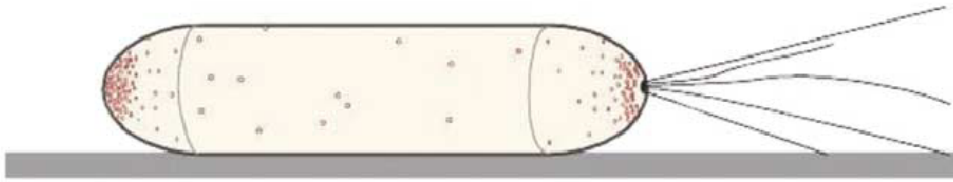


# *Actin-driven motility of lipid vesicles coated with ActA.*

Fluorescently labeled ActA molecules (blue)



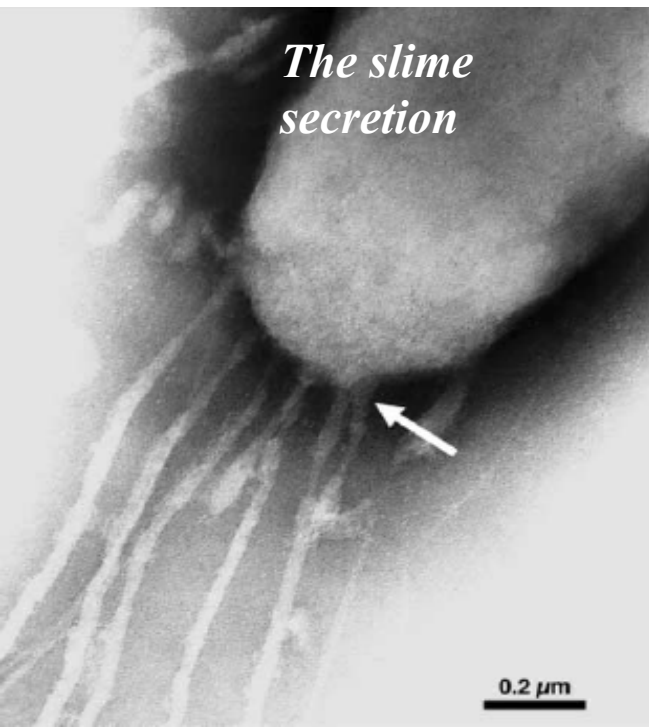
*A moving vesicle deformed by actin.*



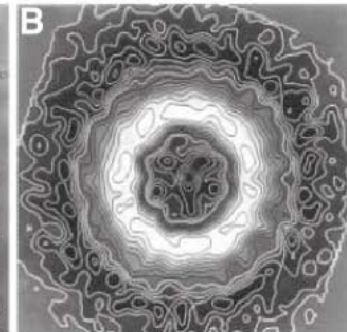
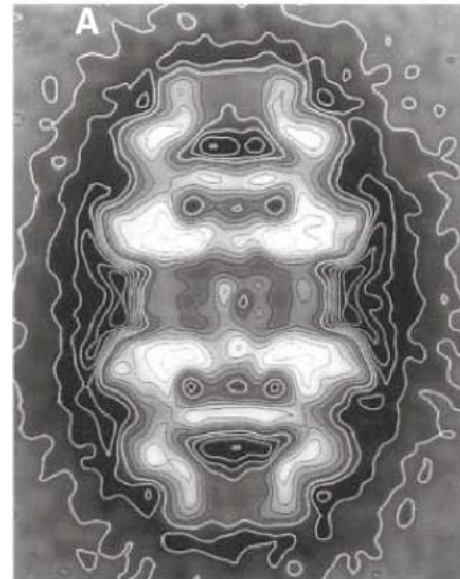
In *M. xanthus* nozzles are clustered at the two cell poles, pili at one pole.

**S motility** is generated by the pili, which extend, attach to nearby cells, and then retract, pulling the cells together.

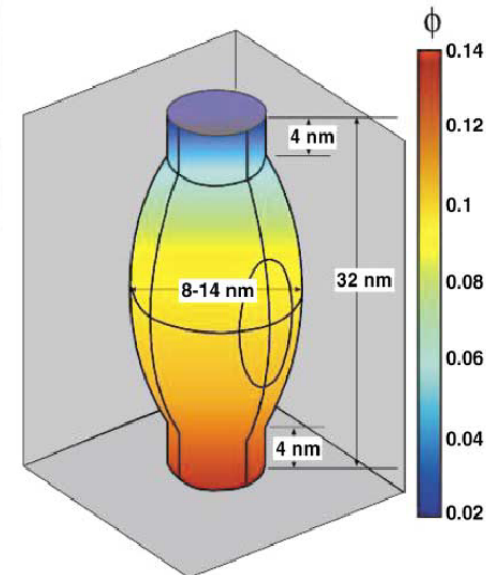
**A motility** is driven by the secretion of mucilage from the nozzles – gliding.



*The slime secretion*



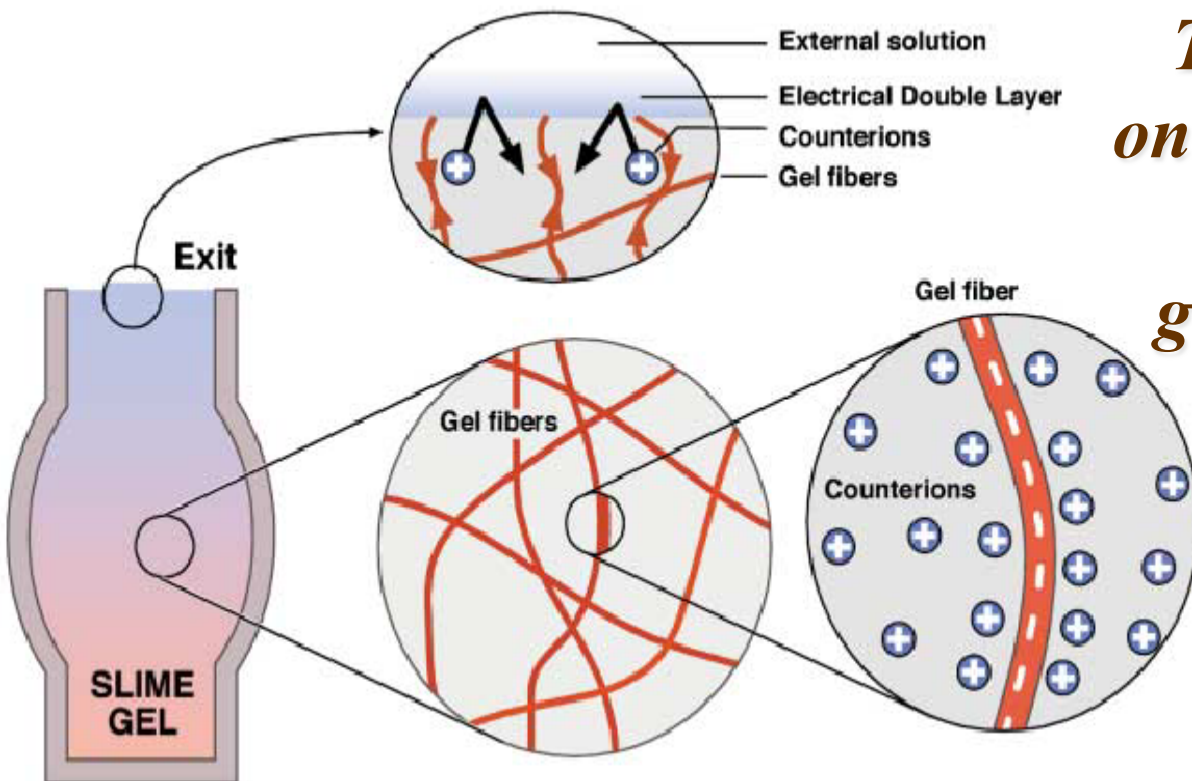
Pore complex in *P. uncinatum*





# *A Model for Nozzle Function*

- + Slime is imported into the proximal end of the nozzle.*
- + Slime is a creoslinked polyelectrolyte gel (crosslinked fibers).*
- + A Donnan potential is generated by the mobile counterions.*
- + The slime is hydrated by water that flows into the nozzle causing the slime to swell.*



*The nozzle hydrates only through the nozzle exit – a hydration gradient exists along the nozzle.*



# Force generation in polyelectrolyte gels

*Negatively charged filaments are surrounded by positive counterions that are confined inside gel by the Donnan potential.*

## *The swelling pressure*

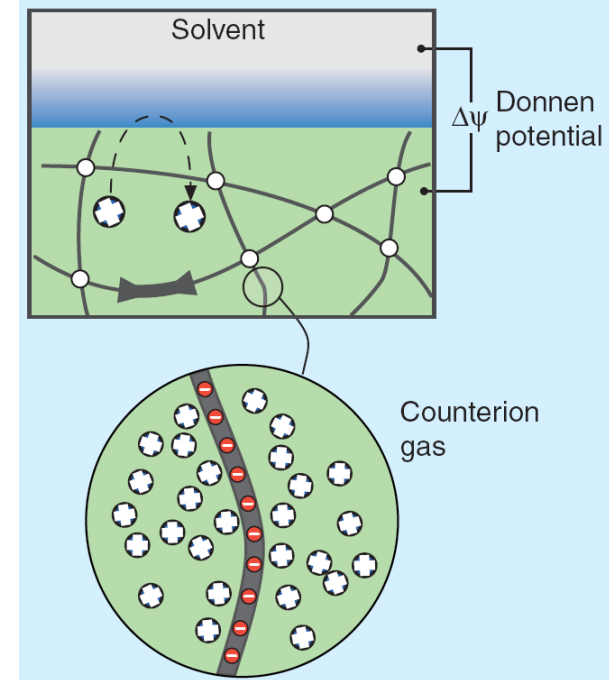
$$\Pi = \Pi_{Entropic} + \Pi_{Ion} + \Pi_{Elastic} + \Pi_{Interactions}$$

$\Pi_{Entropic}$  – the gel fibers tend to diffuse outward.

$\Pi_{Ion}$  – a polyelectrolyte gel contains diffusible counterions. Water will diffuse in.

$\Pi_{Elastic}$  – Gel elasticity tends to resist its tendency to expand outwards.

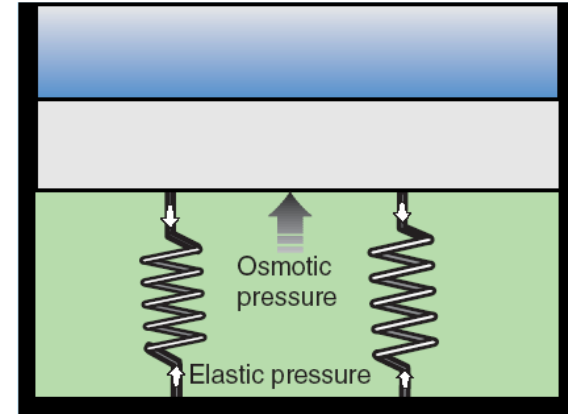
$\Pi_{Interaction}$  – An attraction between the gel fibers.



$$\Pi_{Entropic} + \Pi_{Interactions} \ll \Pi_{Ion} + \Pi_{Elastic}$$

$$\Pi_{swell} = \Pi_{osm} - \Pi_{elas}$$

*At equilibrium, the elastic tension just counterbalances the osmotic pressure of the gel counterions.*



If the cross-links are partially removed – the gel partially ‘solates’ – the elasticity of the gel weakens, allowing the osmotic pressure to expand the gel to a larger volume.

*The force of expansion is in the range of hundreds of pN per square micron, and the expansion would take a few tenths of a second for a micron sized ball.*

